

Testing of 17 sample extracts and procedure blanc for their potential to interfere with TTR-T4 binding using the FITC-T4 assay

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Introduction

In response to a request by Arnika, Prague, Czech Republic, BioDetection Systems b.v. ("BDS") tested the potency of 17 sample extracts, suspected to be contaminated with PFAS, and one procedure blank for their potency to interfere with the binding of the thyroid hormone thyroxine (T4) to the plasma transport protein Transthyretin (TTR) using the FITC-T4 binding bioassay.

Thyroid hormones are important for numerous physiological processes such as regulation of metabolism, bone remodelling, cardiac function, and mental status. Thyroid hormones are of special importance in fetal development, as development of the brain is dependent on normal levels of thyroid hormones. During the first part of pregnancy, the fetus relies entirely on transplacental transfer of maternal thyroid hormones and thus on a normal maternal thyroid function.

Maternal thyroid homeostasis is also contributing to fetal development during the remaining part of pregnancy. Therefore, maintenance of normal thyroid function and levels is essential.

Most of the thyroid hormones are transported in the blood to target organs by the transport proteins TBG, TTR or albumin. Only a small portion of thyroid hormone circulating in the blood is unbound. Unbound thyroid hormone is easily secreted from the body through the urine or faeces. Although the major transport protein is TBG, TTR is of importance since it is involved in crossing the serum-placental and serum-cerebrospinal fluid barriers for the thyroid hormone thyroxine (T4).

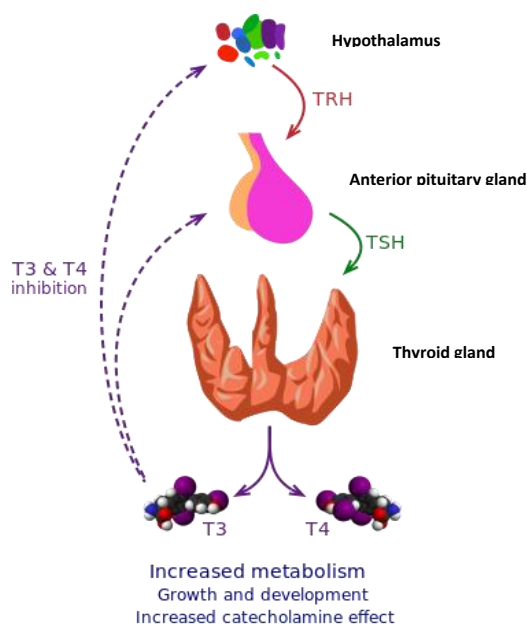


Figure 1 The thyroid system. Regulation of Thyroid hormone production

The thyroid hormone homeostasis can be disrupted by environmental chemicals at different points of interaction in the thyroid pathway, among which during transport of the hormone through the blood. Some chemicals are known to bind to the transport protein TTR thereby replacing the endogenous ligand T4. As a result, T4 is present in the circulating blood unbound and readily excreted. PFAS are such chemicals known for their capability to bind TTR thereby replacing T4.

In the TTR - FITC-T4 binding bioassay, competition between a fixed concentration of FITC-T4 and a dilution series of test items for binding TTR is determined. The measurement is based on the difference in fluorescence between bound and non-bound FITC-T4 to the TTR-binding site. Bound FITC-T4 will result in a higher fluorescence than non-bound.

The presence of increasing concentrations of PFAS, capable of competing with FITC-T4 for TTR-binding sites, will result in a decreased amount of FITC-T4 bound TTR and thereby decreased fluorescence. Disruption of FITC-T4-TTR binding is benchmarked against the reference compound PFOA (potency factor = 1). The amount of fluorescence is proportional to the amount of FITC-T4-specific receptor binding.

In the present study, the potency of 17 migration-sample extracts and procedure blank for disrupting thyroid hormone TTR transport was evaluated using the TTR - FITC-T4 binding bioassay.

This report summarises all analysis results obtained for the received sample extracts.

Sample processing

18 samples including 1 procedure blanc were received from Arnika, Prague, Czech Republic. Upon arrival, all samples were given a unique BDS sample code. In table 1, all sample codes and sample information are given.

Table 1 sample information

| Sample Code | BDS code | UCT Code | Extract weight (g) | Extract volume (mL) |
|------------------------|----------|-----------|--------------------|---------------------|
| Compost-DE-12 | 39752 | LN 24/21 | 1.6282 | 25 |
| DE-PAP-KFC-17a | 39753 | LN 26/21 | 0.3469 | 25 |
| DE-PAP-MCD-26 | 39754 | LN 30/21 | 0.3539 | 25 |
| Compost-DK-3 | 39755 | LN 33/21 | 1.1304 | 25 |
| Compost-DK-5 | 39756 | LN 35/21 | 1.1966 | 25 |
| DK-PAP-MCD-1 | 39757 | LN 38/21 | 0.407 | 25 |
| Compost-NL-3 | 39758 | LN 41/21 | 0.9481 | 25 |
| Compost-NL-1 | 39759 | LN 42/21 | 1.8759 | 25 |
| NL-MCD-01 | 39760 | LN 45/21 | 0.277 | 25 |
| FastF-FR-5 | 39761 | LN 48/21 | 0.3152 | 25 |
| Compost-FR-2 | 39762 | LN 49/21 | 1.2387 | 25 |
| CZ-FCM-MCD-01b | 39763 | LN 52/21 | 0.5611 | 25 |
| Recycl-CZ-1 | 39764 | LN 53/21 | 0.6594 | 25 |
| CZ-FCM-KFC-06 | 39765 | LN 57/21 | 0.2806 | 25 |
| PizzaB-UK-2 | 39766 | LN 59/21 | 1.0998 | 25 |
| FastF-UK-2 | 39767 | LN 62/21 | 0.3054 | 25 |
| FastF-UK-5b | 39928 | LN 131/21 | 0.4963 | 25 |
| Client procedure blanc | 39929 | | | |

The solvent was evaporated under a gentle stream of N₂, until about 10 µL sample was left. This was dissolved in 1 mL of MeOH and transferred to a pre-rinsed bottle containing 100 mL HPLC water. The vial was rinsed twice with 0.5 mL of MeOH and this was also transferred into the pre-rinsed bottle.

A weak anion exchange (WAX) solid phase extraction (SPE) was performed to clean up the samples. The cartridge was conditioned using:

- 4 ml of MeOH with 0.1% NH₄OH
- 4 ml of MeOH
- 4 ml HPLC water

The sample was transferred to the cartridge at a rate of about 1 drop per second. The cartridge was washed using:

- 4 ml 25 mM NH₄Ac pH 4
- 8 ml THF:MeOH (75:25)

After washing, the cartridge was dried for 30 minutes applying vacuum. A 15 ml tube was placed under the cartridge and the sample was eluted using 4 ml of MeOH with 0.1% NH₄OH.

The extract was evaporated to dryness under a gentle stream of N₂ and re-dissolved in 50 µL of DMSO followed by preparation of serial dilutions in DMSO.

TTR - FITC-T4 binding assay

Serial dilutions of the sample extracts (2 μ l) or reference material (2 μ L) were incubated in Tris-buffer (pH 8.0) for 20 minutes in the presence of TTR (0.30 μ M) and FITC-T4 (109 nM). The total incubation volume was 202 μ l. After incubation, the fluorescence was measured using a Berthold luminometer applying an excitation wavelength of 490 nm, an emission wavelength of 535 nm and a measuring time of 0.1s.

Data analysis

Analysis results of the sample extracts expressed as induction relative to the standard reference compound PFOA, are intrapolated in the calibration curve for quantitative determination of thyroid hormone transport disruptive potential using the statistical software package Graphpad Prism V5.03.

Results

In figures 2-5, the TTR - FITC-T4 analysis results of the 17 sample extracts and procedure blanc are presented graphically. In table 2, quantified analysis results are given, expressed as the amount of PFOA equivalents per gram of sample. In addition, the lowest amount of sample showing TTR-T4 disrupting potential per litre of incubation mixture (Lowest Observed Effect Concentration; LOEC) is given.

In the client procedure blanc as well as the laboratory procedure blanc sample, a low signal was measured as reported in Table 2.

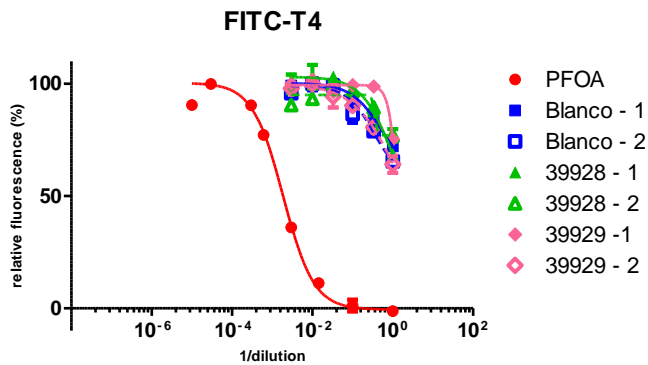


Figure 2

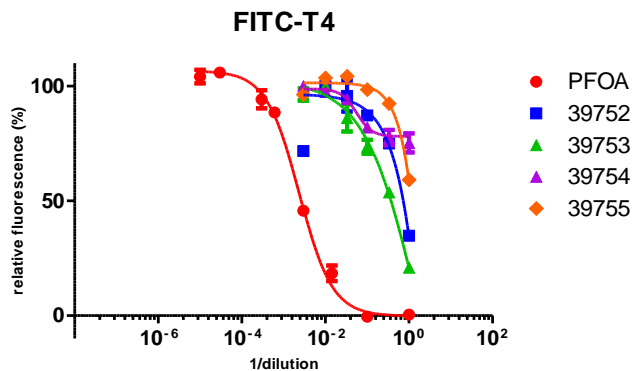


Figure 3

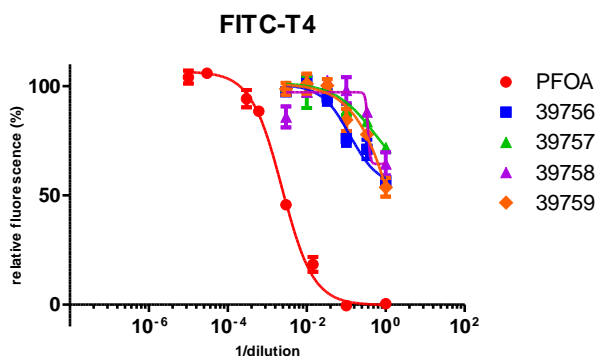


Figure 4

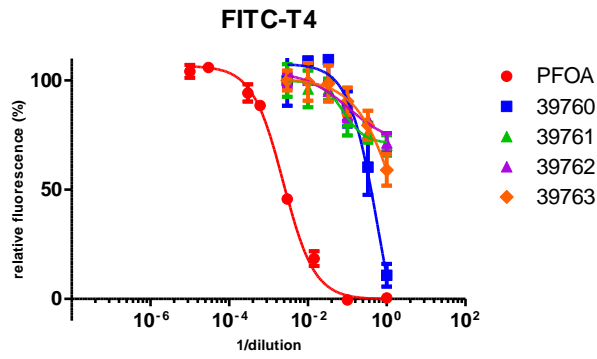


Figure 5

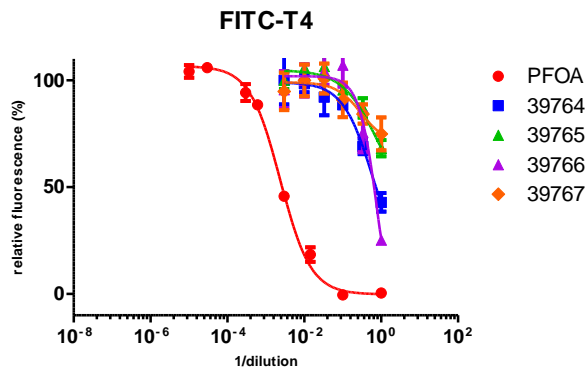


Table 2 TTR - FITC-T4 analysis results

| BDS code | Client code | TTR - FITC-T4 activity (µg PFOA/g sample) | LOQ (µg PFOA/g sample) | LOEC (g sample/l incubate) |
|----------|------------------------|--|---------------------------|-------------------------------|
| Blanc | Laboratory blanc | 21 | 12 | 2.4 |
| 39752 | Compost-DE-12 | 23 | 5.6 | 3.1 |
| 39753 | DE-PAP-KFC-17a | 341 | 26 | 0.2 |
| 39754 | DE-PAP-MCD-26 | 180 | 26 | 0.4 |
| 39755 | Compost-DK-3 | 13 | 8.1 | 5.5 |
| 39756 | Compost-DK-5 | 74 | 7.7 | 1.0 |
| 39757 | DK-PAP-MCD-1 | 51 | 23 | 1.4 |
| 39758 | Compost-NL-3 | 27 | 10 | 2.7 |
| 39759 | Compost-NL-1 | 18 | 4.9 | 4.1 |
| 39760 | NL-MCD-01 | 200 | 33 | 0.4 |
| 39761 | FastF-FR-5 | 220 | 29 | 0.3 |
| 39762 | Compost-FR-2 | 21 | 7.4 | 3.5 |
| 39763 | CZ-FCM-MCD-01b | 52 | 16 | 1.4 |
| 39764 | Recycl-CZ-1 | 73 | 14 | 1.0 |
| 39765 | CZ-FCM-KFC-06 | 69 | 33 | 1.1 |
| 39766 | PizzaB-UK-2 | 26 | 8.3 | 2.9 |
| 39767 | FastF-UK-2 | 60 | 30 | 1.2 |
| 39928 | FastF-UK-5b | 39 | 19 | 1.9 |
| 39929 | Client procedure blanc | 21 | 12 | 3.5 |